Sample Collection and Processing: CSF

Steps 1-3

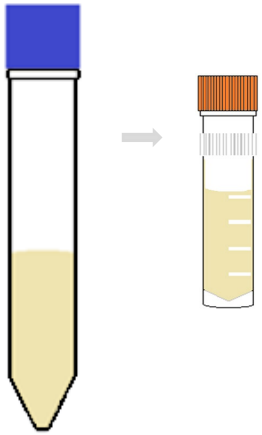
- Prepare Mr. Frosty™

- Chill 15 ml conical tubes in their wrappings on wet ice.
- Label twenty *clear*-capped 2 ml cryotubes with preprinted PBN CSF labels.
- Label one *orange*-capped 2 ml cryotube with CSF PELLET label.
- Label one *orange*-capped 2 ml cryotube with CSF NMDA label.

Steps 4-5

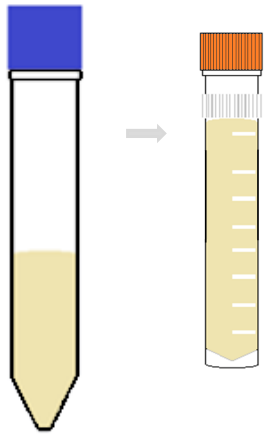
- Collect 15ml of CSF into 1 centrifuge tube.

Step 6



- Dispense 1ml into the CSF NMDA-labelled orange-capped cryovial

Step 7

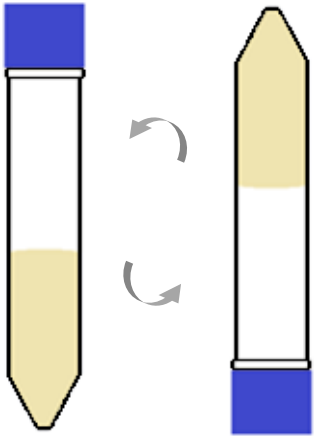


- Dispense 2-4ml CSF into the 4 ml orange-capped cryovial.
- Send to local pathology lab for testing. Label and handle sample per your local path lab's instructions.

Continued on next slide

Sample Collection and Processing: CSF (cont'd)

Step 8



- Gently invert the remaining 10 ml of CSF in the centrifuge tube 3-4 times to mix the sample.

Step 9

- Within 30 minutes of collection, centrifuge samples at 300 x g for 10 minutes at 4° C.
- Aliquot 500 ul of supernatant directly into each of the prepared cryotubes, being careful not to disturb the pellet at the bottom of the conical tube.
- ***Leave 500 ul of CSF in the conical tube.***

Steps 10-12

CSF PELLET

- Add 500 ul CryoStor® to 500 ul of CSF and cell pellet in the 15 ml conical tube.
- Resuspend pellet using pipetting technique.
- Transfer the resuspended CSF pellet to the pre-labeled orange cryotube.
- Within 60 minutes of CSF collection, freeze CSF aliquots **upright** in rack or cryobox at -80° C.
- Place pellet aliquot in the prepared Mr. Frosty™ and store at -80° C overnight.